

PHYTOPLANKTON COMMUNITY STRUCTURE, SEASONAL DYNAMICS AND PHYSICO-CHEMICAL CHARACTERISTICS OF THE RADOINJA RESERVOIR (SERBIA)

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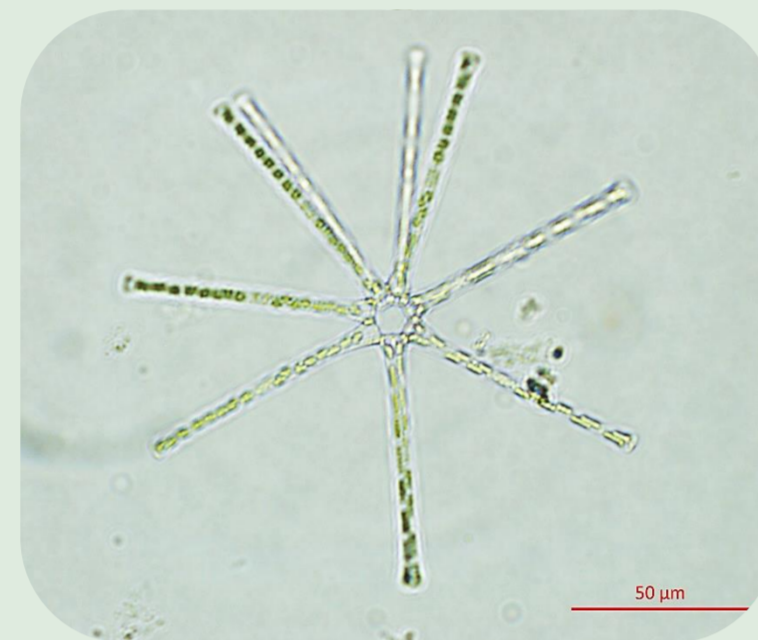
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Introduction

The Radoinja Reservoir is situated in Southwestern Serbia. It represents a water filled canyon meandering between narrow and steep limestone shores and formed by constructing a dam across the Uvac River. The Radoinja Reservoir is 12 km long; it has a total volume of $7.6 \times 10^6 \text{ m}^3$ and useful volume of $4.1 \times 10^6 \text{ m}^3$. The mean altitude of the Radoinja Reservoir is 812 m a.s.l. The Radoinja Reservoir is a multi-purpose; it is used for electricity production and partially for water supply of the town of Priboj and its surrounding.



Cyclotella ocellata Pantocsek



Asterionella formosa Hassall

Materials and Methods

Field research of the Radoinja Reservoir was carried out in June, August and November 2014. The sampling was conducted at 3 sampling sites by depth (near the dam-A₁; central part of the reservoir-B₁ and at the entrance to the reservoir-C₁). By measuring water temperature at every 0.5 m depth, the layers of epilimnion, metalimnion (thermocline) and hypolimnion were determined.

The samples for basic physico-chemical parameters, primary nutrients and chlorophyll *a* were taken at each 1.5 m in epilimnion, at each 0.5 m in metalimnion, and then at every 1.5 m in hypolimnion to the depth of 15 m, and then at each 5 m (including 10% of the depth of bottom of the reservoir). Analysis of physico-chemical parameters was conducted using standard analytical procedures according to appropriate SRPS ISO methods.

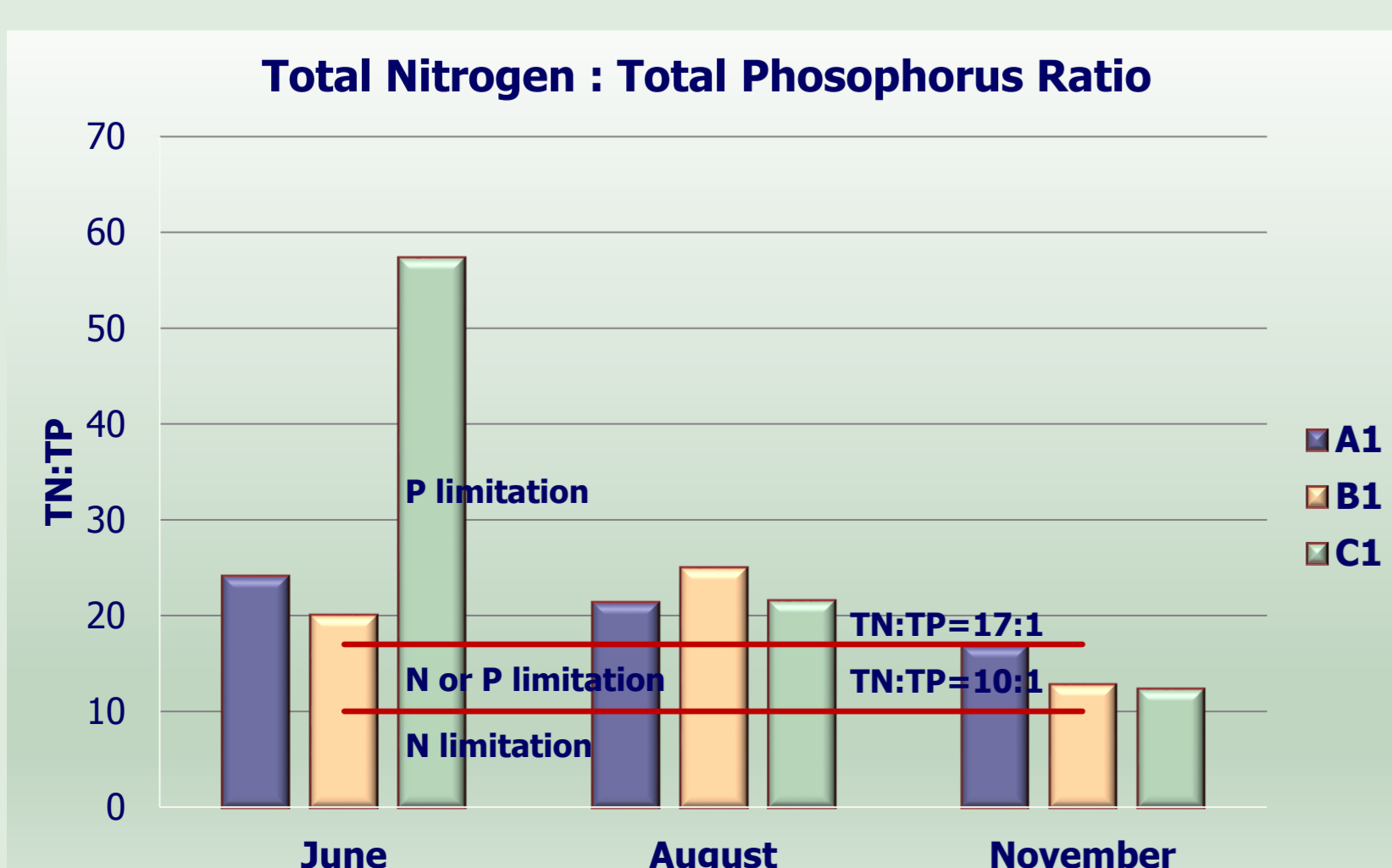
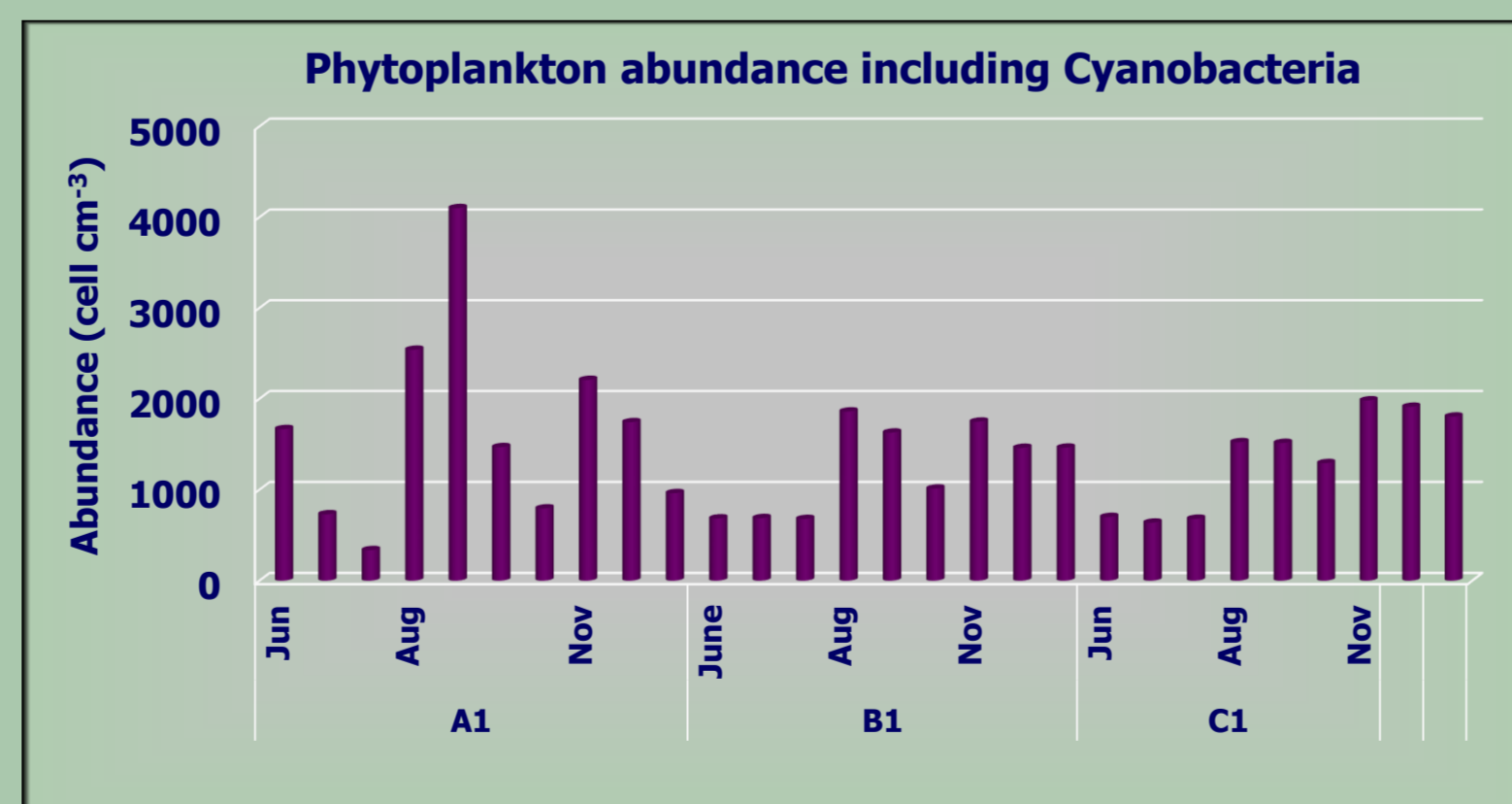
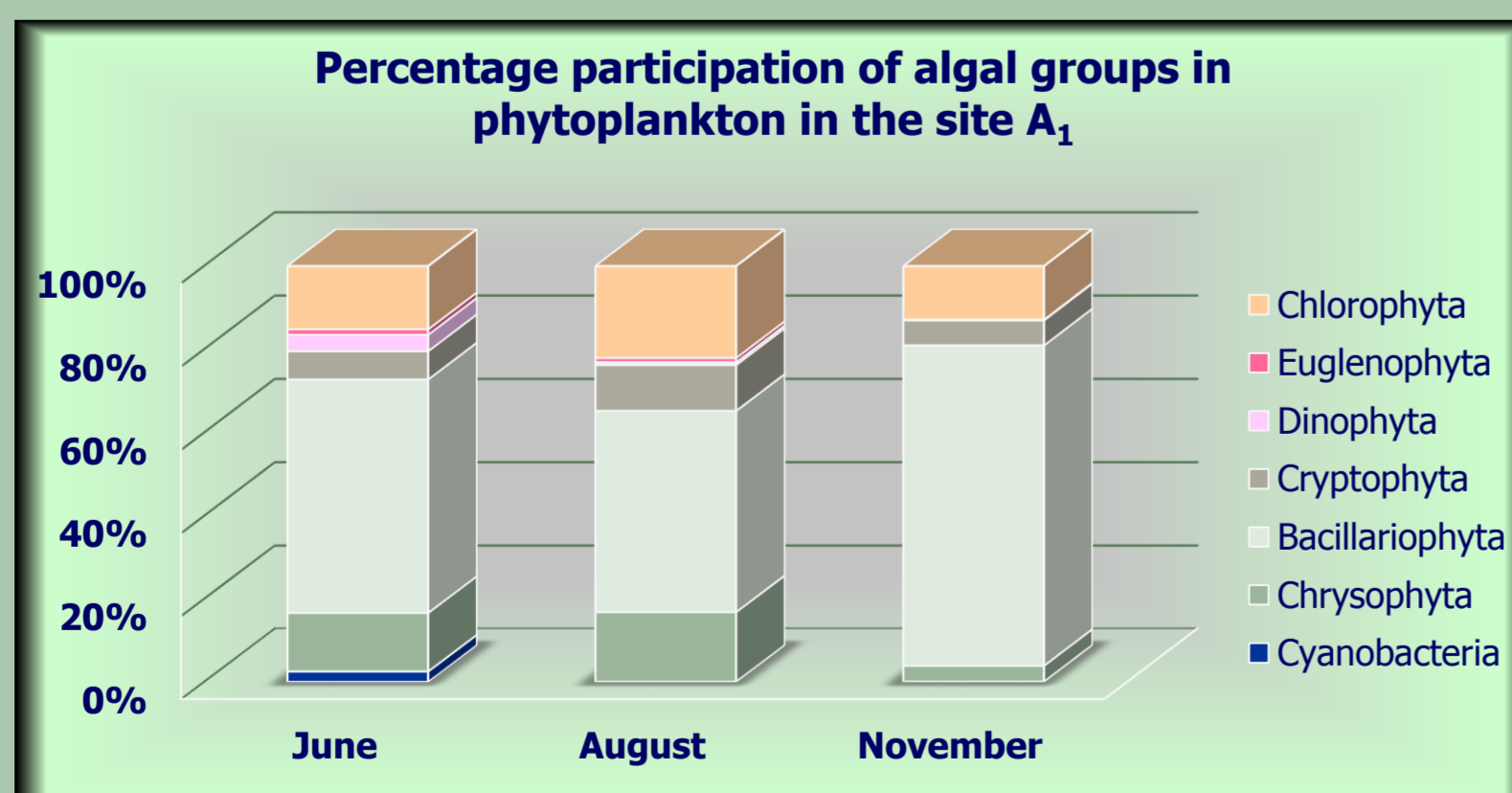
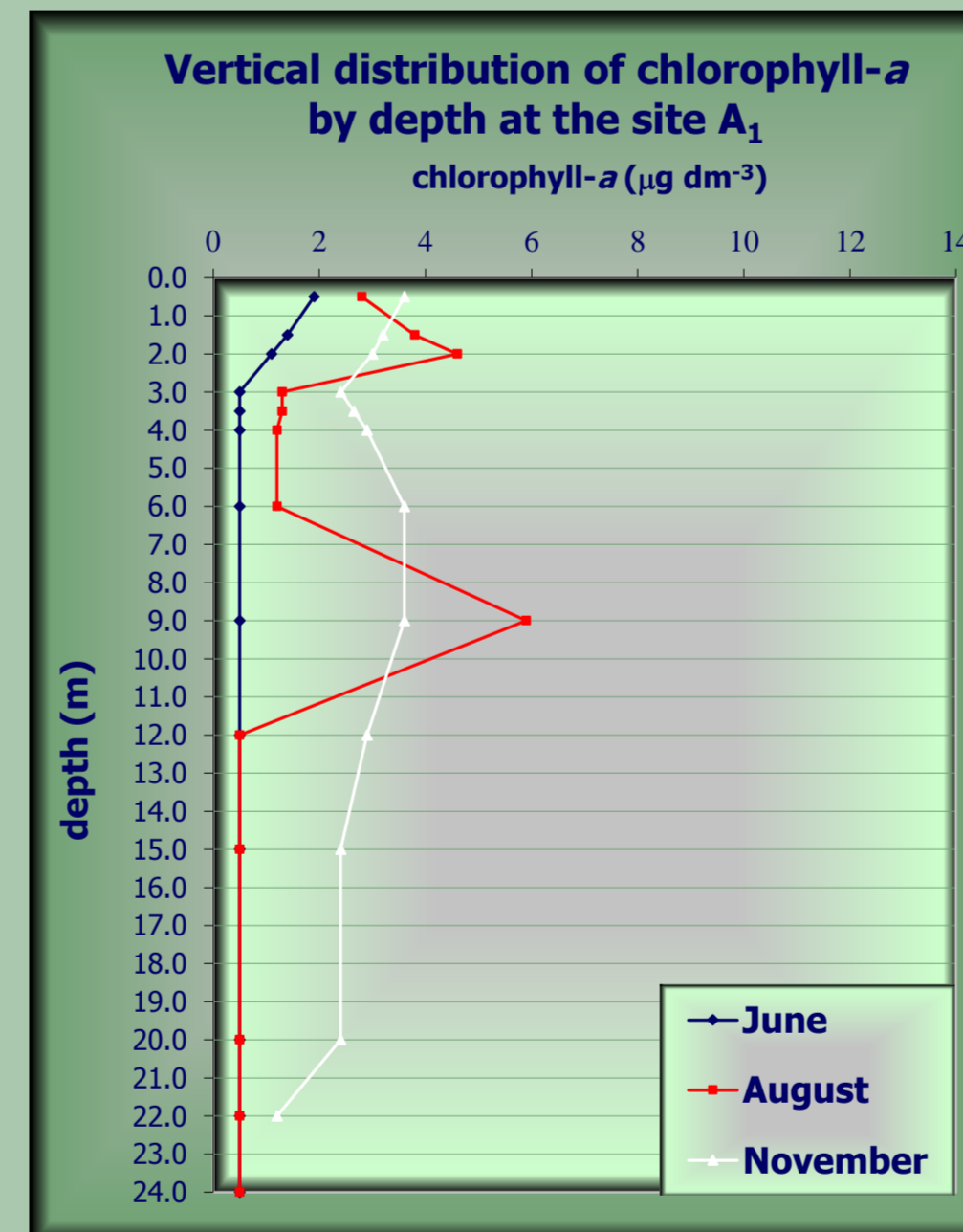
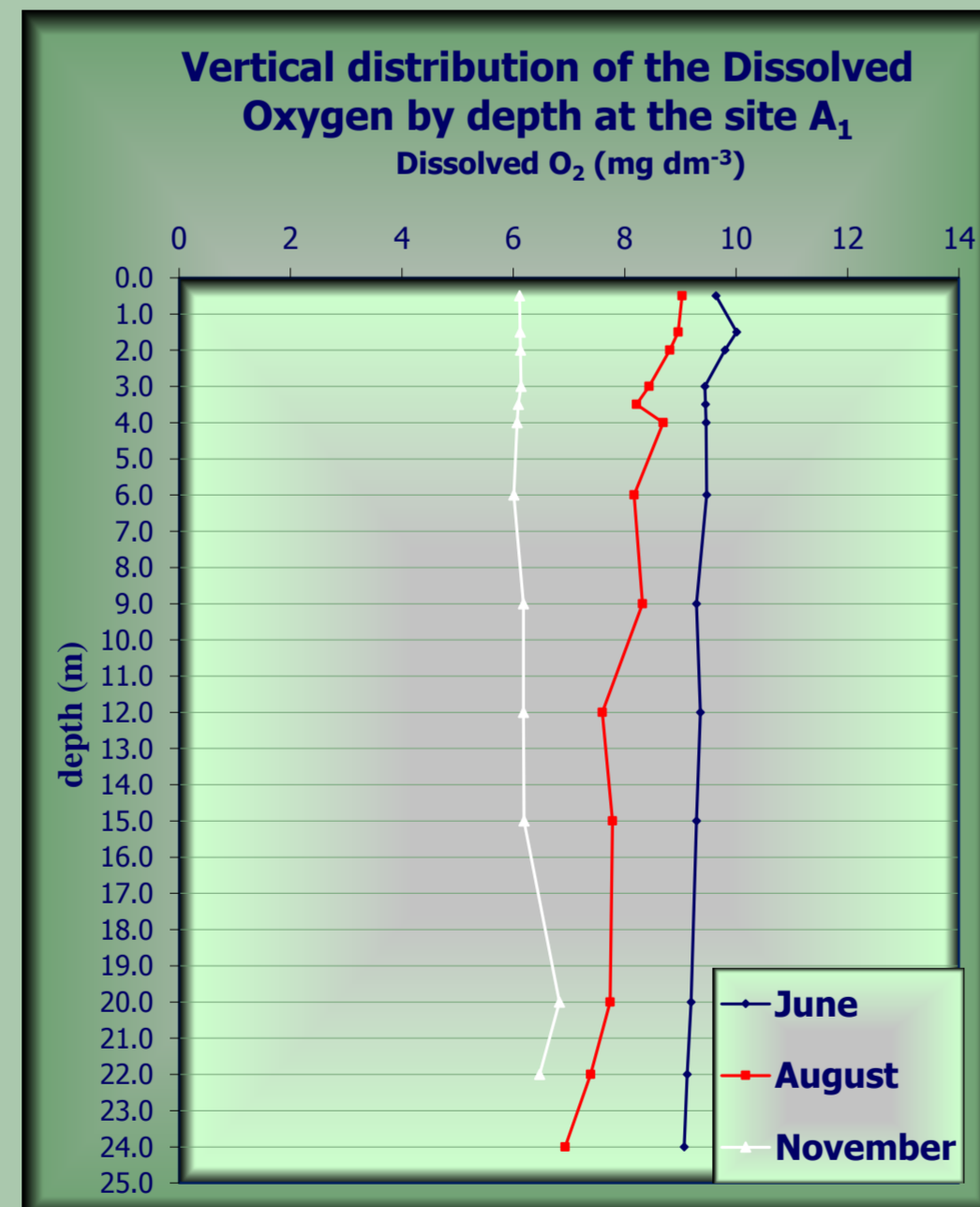
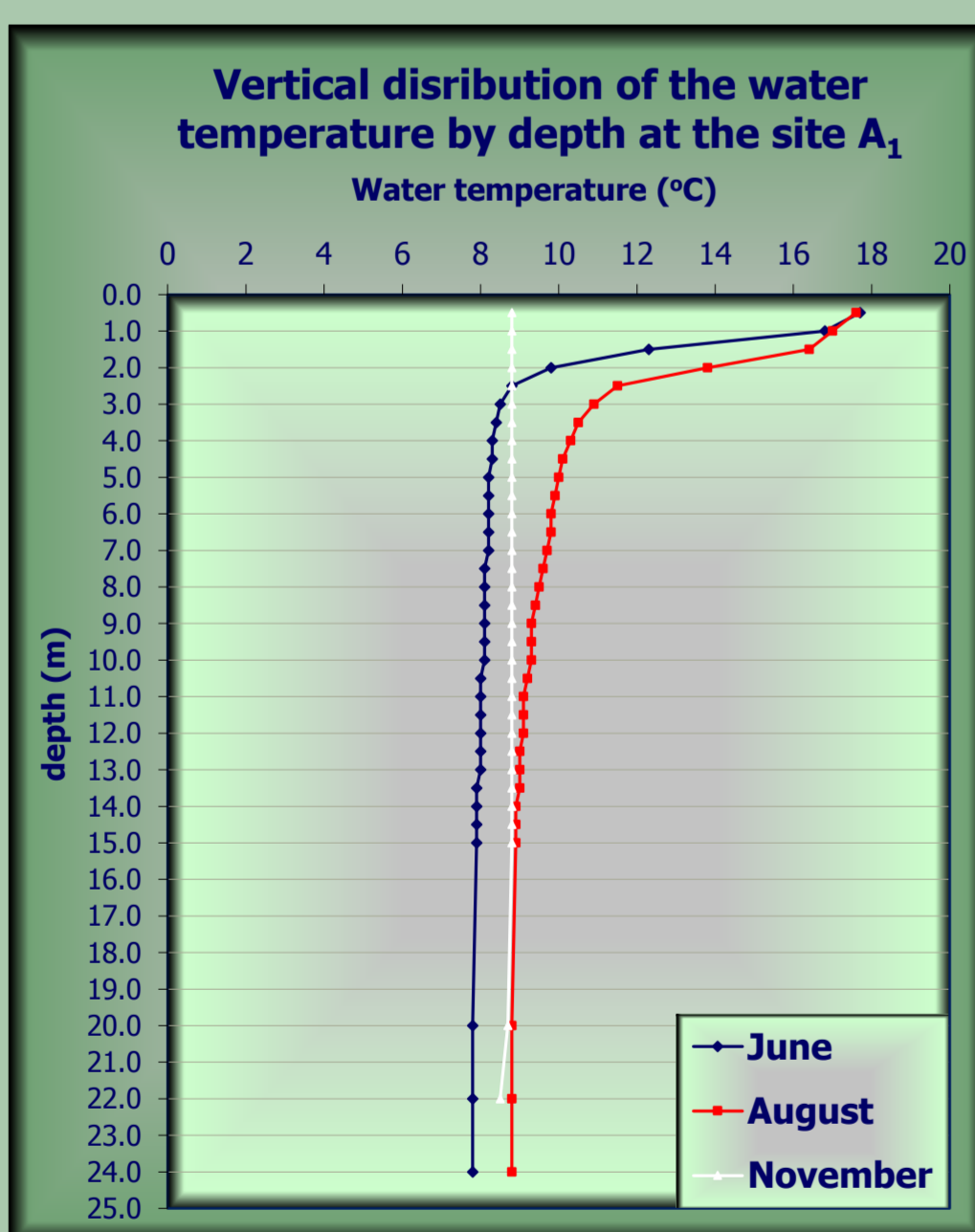
The algal material was collected using plankton nets (25 μm mesh size) and hydrobiological bottles. At three or four sampling sites by depth, two samples for quantitative analysis of phytoplankton were taken, and preserved with formaldehyde or Lugol's iodine. The phytoplankton analysis was done on inverted microscopes: Nikon TE-2000U with DS-5M camera and NIS-Elements D software and Zeiss Axiovert with AxioCam HRC camera and AxioVision 4.8 software. Quantitative analysis of phytoplankton was done using Utermöhl method (1958) according to the SRPS EN 15204: 2008.

Results and Discussion

The first two field research is characterised by thermal stratification whilst the third by total circulation of the reservoir. During all year the Dissolved Oxygen (DO) concentration was high in all water layers; there was not the DO deficit in the hypolimnion. The investigation showed that the phosphorus concentration is a limit factor for phytoplankton growth due to the TP:TN ratio > 17:1 and the Dissolved Inorganic Nitrogen (DIN):Soluble Reactive Phosphorus (SRP) mass ratio > 10:1. By qualitative phytoplankton analysis a total of 117 taxa were identified from 6 algal divisions: Chrysophyta (3 taxa), Bacillariophyta (52 taxa), Dinophyta (5 taxa), Cryptophyta (3 taxa), Euglenophyta (6 taxa), Chlorophyta (38 taxa) and Cyanobacteria (10 taxa). Quantitatively, the diatoms were dominant. Besides typical planktonic forms, the benthic diatoms were numerous. The highest phytoplankton abundance was recorded in August 2014, at the deepest sampling point, near the dam at 2.0 m depth, and it was $4100 \text{ cell cm}^{-3}$ whilst the lowest phytoplankton abundance was in June 2014, near the dam, too, at the 24 m depth (336 cell cm^{-3}). The dominant species in phytoplankton community was *Cyclotella ocellata*. The Radoinja Reservoir belongs to oligo-mesotrophic reservoirs characterised by relatively uniform phytoplankton community composition and low productivity as well. In the summer period, a maximum of phytoplankton growth was observed, with poorly expressed peak below the thermocline (the highest chlorophyll *a* concentration was at 9.0 m depth and it was $5,9 \mu\text{g dm}^{-3}$). Due to low phytoplankton productivity a water transparency was high (over 7 meters).

Phytoplankton taxa list of the Radoinja Reservoir in 2014

Cyanobacteria	Rhodospaenia abbreviata (C. Agardh) Lange-Bertalot
<i>Aphanocapsa incerta</i> (Lemm.) Cronberg & Kom.	<i>Rhopalodia gibba</i> (Ehrenberg) Otto Müller
<i>Cuspidothrix issatschenko</i> (Usachev) P.Rajaniemi, Komárek, R.Willame, P. Hrouzek, K.Kastovská	<i>Staurisira construens</i> Ehrenberg
<i>Gleotrichia echinulata</i> P.G. Rihter	<i>Staurisira venter</i> (Ehrenberg) Cleve & J.D. Müller
<i>Kamptomena formosum</i> (Bory de Saint-Vincent ex Gomont) Struněký, Komárek & J. Smarda	<i>Stephanodiscus minutulus</i> (Kützting) Cleve & Möller
<i>Merismopedia elegans</i> A. Braun in Kützting	<i>Suriella brebissonii</i> Krammer & Lange-Bertalot
<i>Oscillatoria limosa</i> Agardh ex Gomont	Cryptophyta
<i>Phormidium terebriforme</i> (Agardh & Gom.) Anagnostidis & Komárek	<i>Cryptomonas</i> Ehrenberg sp.
<i>Planktithrix rubescens</i> (DeCand. ex Gom.) Anagn. & Kom.	<i>Plagiocelmis nanoplantica</i> (H.Skuja) G.Novario, I.A.N. Lucas & S.Morall
<i>Pseudonabaena limnetica</i> (Lemm.) Kom.	<i>Rhodomonas lacustris</i> Pascher & Ruttner
<i>Snowella lacustris</i> (Chodat) Kom. & Hindák	Dinophyta
Chrysophyta	<i>Ceratium hirundinella</i> (O.F. Müller) Dujardin
<i>Chrysoococcus biporus</i> Skuja	<i>Gymnodinium</i> Stein sp.
<i>Dinobryon divergens</i> Imhof	<i>Parvodinium incognitum</i> (Lemmermann) S. Carty
<i>Dinobryon sertularia</i> Ehrenberg	<i>Peridinium</i> Ehrenberg sp.
Bacillariophyta	<i>Peridinium cinctum</i> Ehrenberg
<i>Achnantheidium catenatum</i> Bily & Marvan	Euglenophyta
<i>Achnantheidium minutissimum</i> Kützting	<i>Euglena</i> Ehrenberg sp.
<i>Amphora ovalis</i> (Kützting) Kützting	<i>Lepidocisilus acis</i> (O.F. Müller) Marin & Melkonian
<i>Amphora pediculus</i> (Kützting) Grunow	<i>Monorophina pyriformis</i> (Ehrenberg) Mereschkowsky
<i>Asterionella formosa</i> Hassall	<i>Strombomonas acuminata</i> (Schmarda) Deflandre
<i>Campylodiscus hibernicus</i> (Ehrenberg)	<i>Trachelomonas hispida</i> (Perty) Stein em. Deflandre
<i>Caloneis silicula</i> (Ehrenberg) Cleve	<i>Trachelomonas volvocina</i> Ehrenberg
<i>Coconeis placentula</i> Ehrenberg	Chlorophyta
<i>Cyclotella ocellata</i> Pantocsek	<i>Acutodesmus acuminatus</i> (Lagerheim) Tsarenko
<i>Cymatopleura sola</i> (Brébisson) W. Smith	<i>Acutodesmus obtusius</i> (Turpin) Hegewald & Hanagata
<i>Cyclotella radiosa</i> (Ehrenberg) Kützting	<i>Trachelomonas volvocina</i> Ehrenberg
<i>Cyclotella</i> (Kützting) Brébisson sp.	<i>Chlamydomonas</i> Ehrenberg sp.
<i>Cymbella cistula</i> (Ehrenberg) Kirchner	<i>Chlorococcales</i> sp.
<i>Cymbella affinis</i> Kützting	<i>Closterium aciculare</i> West
<i>Cymbella</i> C. Agardh sp.	<i>Closterium acutum</i> Bréb. var. <i>variabile</i> (Lemm.) Krieger
<i>Denticula tenuis</i> Kützting	<i>Coelastrum astroaleum</i> De Notaris
<i>Diatoma ehrenbergii</i> Kützting	<i>Coelastrum microporum</i> Nägeli
<i>Diatoma moniliformis</i> Kützting	<i>Cosmarium</i> Corda ex Ralfs sp.
<i>Diatoma vulgare</i> Bory	<i>Cosmarium depressum</i> (Nägeli) Lundell var. <i>planctonicum</i> Reverdin
<i>Diplois elliptica</i> (Kützting) Cleve	<i>Crucigenia tetrapedia</i> (Kirch.) W. West & G. S. West
<i>Encyonema minutum</i> (Hilse) D.G. Mann	<i>Desmodesmus bicaudatus</i> (Deslousens) P.M. Tsarenko
<i>Encyonema silicicium</i> (Bleisch) D.G. Mann	<i>Desmodesmus opollensis</i> (P.G. Richter) E. Hegewald
<i>Encyonopsis subminuta</i> Krammer & E. Reichardt	<i>Golenkia radiata</i> Chodat
<i>Ellerbeckia arenaria</i> (Moore & Ralfs) R.M. Crawford	<i>Harlotina reticulata</i> P.A. Dangeard
<i>Fragilaria acuta</i> sensu Krammer & Lange-Bertalot	<i>Hindakia tetrachotoma</i> (Printz) C. Bock, Proschold & Krientez
<i>Fragilaria dilatata</i> (Brébisson) Lange-Bertalot	<i>Hyaloraphidium contortum</i> Pascher
<i>Fragilaria capucina</i> Desmazières	<i>Kirchneriella lunaris</i> (Kirchn.) Moeb.
<i>Fragilaria crotonensis</i> Kitton	<i>Koilella planctonica</i> Hindák
<i>Fragilaria acuta</i> sensu Krammer & Lange-Bertalot	<i>Lagerheimia elliptica</i> (Lagerh.) Chodat
<i>Fragilaria luna</i> sensu Krammer & Lange-Bertalot	<i>Lagerheimia generensis</i> (Chodat) Chodat
<i>Gomphonema olivaceum</i> (Hornemann) Kützting	<i>Monactinus simplex</i> (Meyen) Corda
<i>Gomphonema parvulum</i> (Kützting) Kützting	<i>Monactinus simplex</i> var. <i>echinulatum</i> (Wittrock) Pérez, Maidana & Comas
<i>Gomphonema pumilum</i> (Grunow) E. Reichardt & Lange-Bertalot	<i>Monoraphidium contortum</i> (Thurs.) Komarkova-Legn.
<i>Gyrodinium acuminatum</i> (Kützting) Rabenhorst	<i>Monoraphidium griffithii</i> (Berkeley) Komarkova-Legn.
<i>Gyrodinium attenuatum</i> (Kützting) Rabenhorst	<i>Monoraphidium komarkovae</i> Nygaard
<i>Melosira varians</i> Agardh	<i>Oocystis lacustris</i> Chodat
<i>Navicula</i> Bory de St. Vincent sp.	<i>Pandorina morum</i> (O.F. Müller) Bory
<i>Navicula capitatoradiata</i> Germain	<i>Pleurosigma duplex</i> Meyen
<i>Navicula cryptotenella</i> Lange-Bertalot	<i>Pseudopeleastrum boryanum</i> (Turpin) E. Hegewald
<i>Navicula gregaria</i> Donkin	<i>Scenedesmus grahnseii</i> (Heynig) Fott
<i>Navicula lanceolata</i> (Agardh) Ehrenberg	<i>Scenedesmus quadricauda</i> (Turp.) Brébisson
<i>Navicula radiosa</i> Kützting	<i>Staurastrum chaetoceros</i> (Schroder) G. M. Smith
<i>Nitzschia</i> Hassall sp.	<i>Staurastrum gracile</i> Ralfs ex Ralfs
<i>Nitzschia denticula</i> Grunow	<i>Tetraedron minimum</i> (A. Braun) Hansgirg
<i>Pleurosigma laevis</i> (Ehrenberg) Kom.	<i>Tetrastrum staurigenaeforme</i> (Schroder) Lemm.



Conclusion

In summer period in oligotrophic lakes the available primary nutrient content which controls productivity is suddenly decreased in surface water layer "suppressing" the phytoplankton populations to some kind of a refuge in the zone below thermocline. In surface water layer only a small amount of renewable primary nutrients remains enabling the survival of poor summer plankton community. Such conditions were noted in the Radoinja Reservoir, which belongs to oligo-mesotrophic type, and its phytoplankton community was characterised by uniform composition and low productivity.

